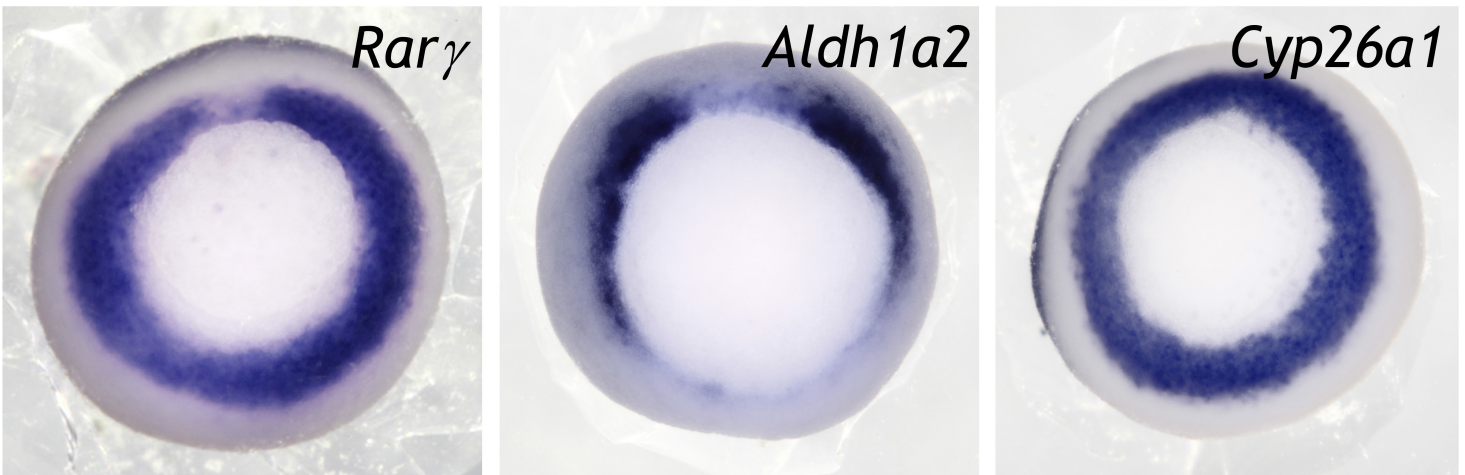


Supplemental Figure S1. *RAR γ 1* is required for the expression of mesoderm markers. Embryos were microinjected unilaterally at the 2-cell or 4-cell stage with 10 ng *Rar γ 1*.L/S MO (see Fig S3). Injected side is to the right of the dotted line, and is indicated by the turquoise β -gal lineage tracer. *Rar γ 1* MO results in the loss of *T (Brachyury)* and *Foxa4*.



Supplemental Figure S2. Whole mount in situ hybridization showing the expression of *Rarγ*, *Aldh1a2*, and *Cyp26a1* at stage 10.5/11, vegetal view, dorsal lip at the top.

>X. laevis Chromosome 2.S

tcttccacctaaggtttataatggatttatcgctgaataactaaggaagacattg
gtaccaaggaagattccaagacaactactggctagcactaagtaagagactgag
c**tagaacaaggca (ATG) GCAACAGC**AGCAAGGAGCGCCTCTGTG

>X. laevis Chromosome 2.L

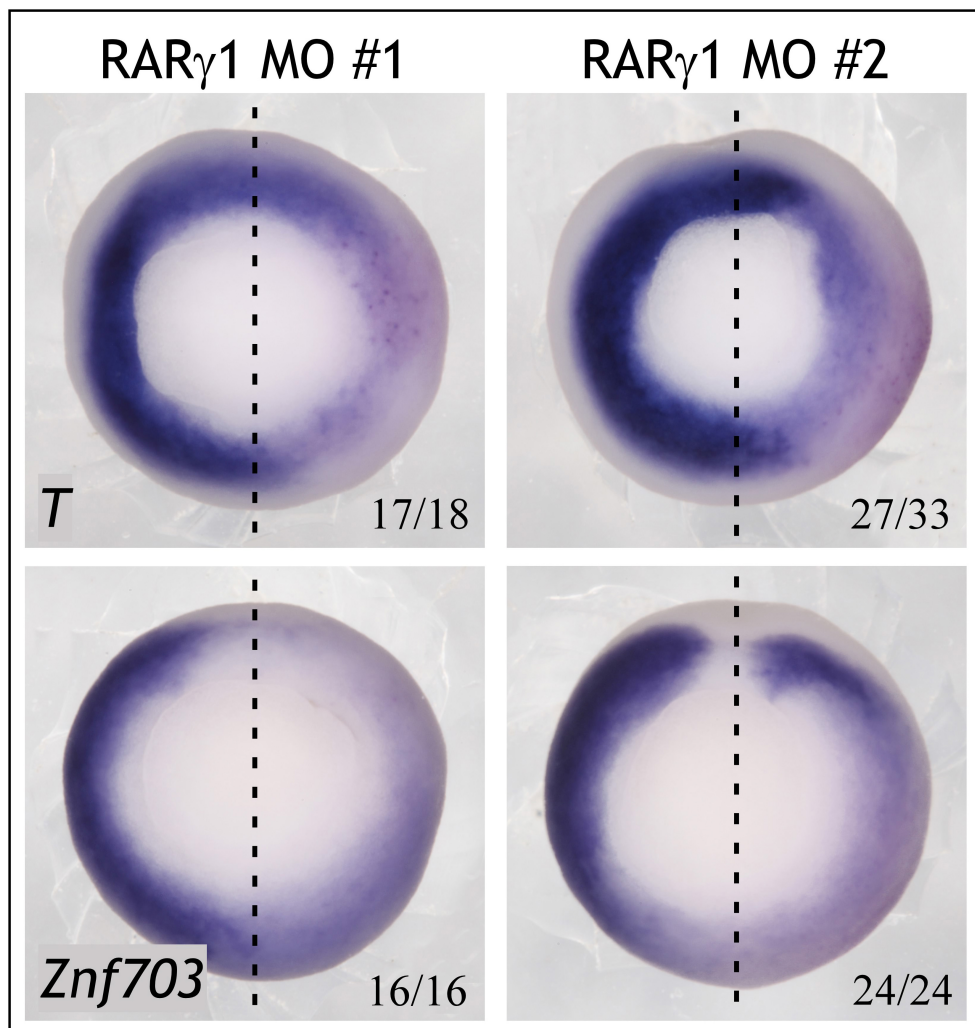
tcttctacctaaggtgtatcatggatttattgctgaataactaaggaagactttg
gtacccaaagaagattccagga-aactactggctagaactcagtgaagagactgag
gg**agaacaaggca (ATG) GCAACAGC**AGCAAGGAGCGCCTCTGTG



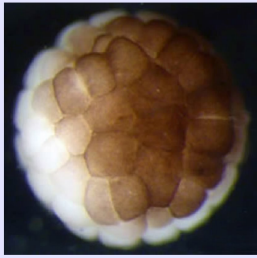
RAR γ 1 MO #1



RAR γ 1 MO #2



Supplemental Figure S3. Specificity of RAR γ 1 MO phenotype. Two different morpholinos were designed to target *Xenopus laevis* Rar γ 1. **(Top)** Mapping of MOs to the .S and .L subgenomes (Session, 2016). MO#1 matches nearly perfectly to both .S and .L, whereas MO #2 matches only .S, and will not likely to bind .L. **(Bottom)** Embryos were microinjected unilaterally at the 2-cell or 4-cell stage with 6.6 ng of either Rar γ 1.L/S MO (#1) or Rar γ 1.S (#2). Injected side is to the right of the dotted line, and is indicated by the magenta β -gal lineage tracer. RAR γ 1 MO #1 and MO#2 give the same knockdown phenotype on *T* and *Znfx703*. Fractions represent the portion of embryos displaying the phenotype.

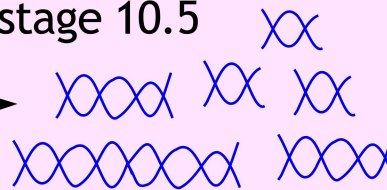
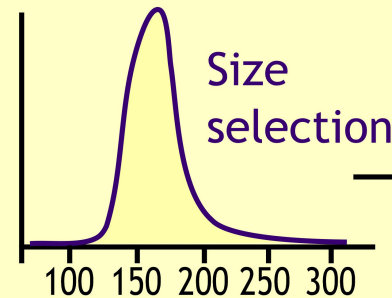
Harvest blastula stage *Xenopus laevis* embryos

Soak with either . . .

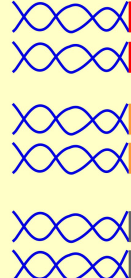


1. 1 μ M RAR-selective agonist TTNPB
2. 1 μ M RAR-selective antagonist AGN193109
3. Vehicle Control (0.1% Ethanol)

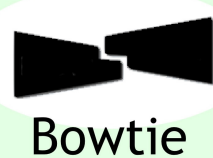
Homogenize embryos at gastrula stage 10.5

cDNA conversion,
fragmentation,
and end repair

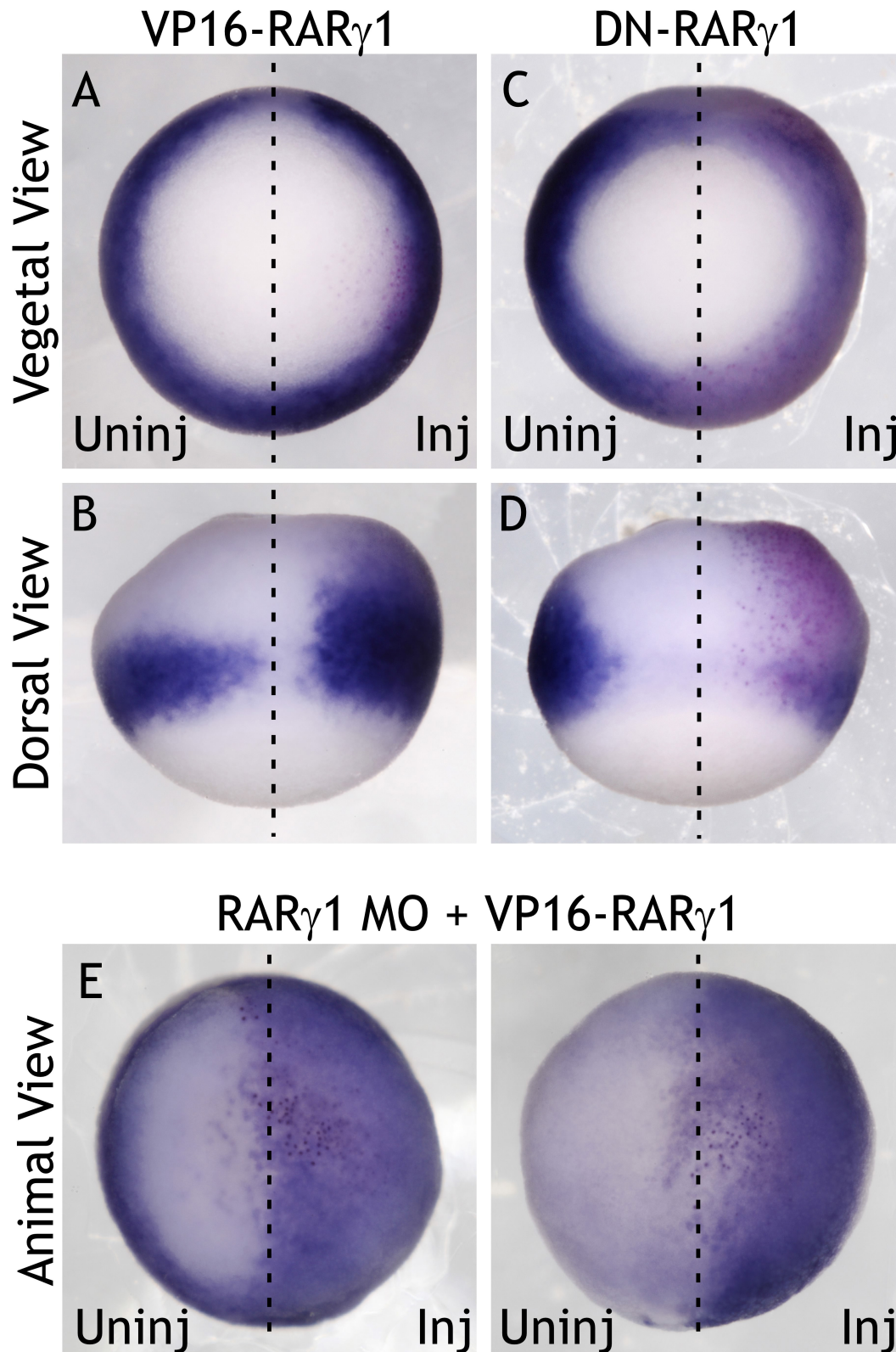
BARCODING

1 μ M TTN1 μ M 109

Vehicle

X. laevis
genome
assembly
Ver. 9.1

Supplemental Figure S4. Schematic of the experimental design and RNA-Seq pipeline. Total RNA was isolated from gastrula (stage 10.5) embryos that had been treated at the early blastula (stage 6) with control vehicle (0.1% ethanol), RAR-selective agonist TTNPB (TTN), or RAR-selective antagonist AGN193109 (109). After cDNA synthesis, multiplexed sequencing reactions were run (Aceto et al., 2014). Reads were mapped to *X. laevis* genome v9.1 (Session et al., 2016) using Bowtie, TopHat and Cufflinks algorithms.



Supplemental Figure S5. VP16-*Rarγ1* mRNA expands *Znf703* while DN-*Rarγ1* mRNA diminishes *Znf703*. (A-E) Embryos were injected unilaterally at 2- or 4-cell stage. Injected side is to the right of the dotted line, and is indicated by magenta β -gal lineage tracer. (A, B, E) 0.2 ng VP16-*Rarγ1* mRNA expands *Znf703* dorsally and animally. (C, D) 2 ng DN-*Rarγ1* mRNA diminishes *Znf703*. Embryos are shown at stage 10.5/11 in vegetal (A, C), dorsal (B, D) or animal (E) view.

Supplemental Table S1 (1 μ M TTNPB) – see Excel sheet

Supplemental Table S2 (1 μ M AGN193109) – see Excel sheet

[Click here to Download Tables S1 & S2](#)

Supplemental Table S3 (Morpholinos)

MO	Sequence (5'→3')
<i>Rara2.L</i>	ATC CAA AGG AAG GTG AGT GTG TGT G
<i>Rara2.S</i>	CTG AAA TCC AAA CTG ACC ATA GAG T
<i>Rary1.L/S</i>	GCT GTT TGC CAT TGC CTT GTT CTA
<i>Rary1.S</i>	CTA GCC AGT AGT TGT CTT GGA ATC T

Supplemental Table S4 (Probe Design)

Probes with T7 Adapters

Primer	Sequence (5'→3')
Aldh1a2_Probe_For	ACCCTTGAATCTCTAAACAGTGGC
Aldh1a2_Probe_Rev	taatacgactcactatagggAATCTCTTCTCTGGCAATCCGCA
Cyp26a1_Probe_For	GCAGGTTCTCTCCAAGTGAAGC
Cyp26a1_Probe_Rev	taatacgactcactatagggCCGCAGAGTCTCCTTAATGACAC
T_Probe_For	GAACGTACAGTACCGGGTGGG
T_Probe_Rev	taatacgactcactatagggTGGTGTGATGGCACTGTTACTC
Gdf3_Probe_For	CTCAGTCTTTCCGTCTCCTTCAC
Gdf3_Probe_Rev	taatacgactcactatagggCAACCACACTCATCCACTACCA
Wnt8_Probe_For	CAACTCTTCTGATCTTCTGCCCA
Wnt8_Probe_Rev	taatacgactcactatagggGTGATTGCCAATATCCCGAAACTC
Skida1_Probe_For	CTGGAGTCGGGCTATGAAGTGG
Skida1_Probe_Rev	taatacgactcactatagggTCCTCCTGTGCCTGTAAGTGG
Kielin_Probe_For	ATTCTGTTGCCACTTCTCTTCTCC
Kielin_Probe_Rev	taatacgactcactatagggATCTCTTCATCCTCCATTTGACGC
Kremen2_Probe_For	TGTTGGTGGAGATGCTGTGG
Kremen2_Probe_Rev	taatacgactcactatagggTCTGGAATCTGGAAGATGTGGA
Nkx6.2_Probe_For	GCCGAGATGAAGACCACTCTG
Nkx6.2_Probe_Rev	taatacgactcactatagggCTCCTCGTTGTCTGAACTCTCC
Mamdc2_Probe_For	ACAGGAAGGGATGTTCTTTGATGC
Mamdc2_Probe_Rev	taatacgactcactatagggTGGCTTTCTGTCCAAACACCA
Znf703_Probe_For	AGCTGAATTCTGTGACCTCCAG
Znf703_Probe_Rev	taatacgactcactatagggCATAAAGCCGTAGGTGTACAAGG
Znf503_Probe_For	CCACTGGGTTCTGGAAGTCCG
Znf503_Probe_Rev	taatacgactcactatagggTTTATAGGGTGACACAGGTGC
Dhh_Probe_For	TGCCTAATGTGCCAGAGAAGACTC
Dhh_Probe_Rev	taatacgactcactatagggACAGATGATTGGGTGTAACAAGGA
Btg2_Probe_For	CTCCCGAAAGTCAGCAAGACAC
Btg2_Probe_Rev	taatacgactcactatagggAGACACAAATTGAACCGTCCTCTC
Hapln3_Probe_For	CTTGCCGAATTTGCTGTGATTCTC
Hapln3_Probe_Rev	taatacgactcactatagggGTTCTCCGTAGCTTCTAACACCA
Fstl1_Probe_For	GAGCCCAAGAGCAAGTCTAAGG
Fstl1_Probe_Rev	taatacgactcactatagggCAGATGGTTGGAAGTCGGGA

Supplemental Table S5 (QPCR)

Primer	Sequence (5'→3')
F (<i>Histone H4</i>):	GAT AAC ATC CAG GGC ATC AC
R (<i>Histone H4</i>):	TAA CCT CCG AAT CCG TAC AG